

Optimal transport-based machine learning to match specific expression patterns in omics data

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Huntington's disease (HD)

- HD is a progressive brain disorder that causes uncontrolled movements, emotional problems, and loss of thinking ability (cognition).
- HD is caused by the HTT gene's mutation.
 - In normal people, the HTT gene contains a triple CAG repeat about 10-35 times.
 - In people with HD, this repeat goes on for 36 or more times.

Onset of disease occurs earlier and deterioration is faster with higher number of CAG repeat.

- HD leads to neuronal cell death.

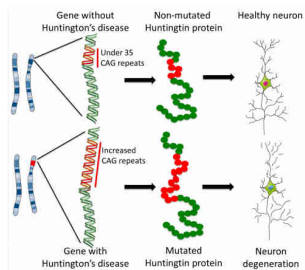


Figure: CAG repeat expansions in HD. Source: California's Stem Cell Agency.

Micro RNAs (miRNAs) and messenger RNAs (mRNAs)

- mRNAs are necessary for translating the genetic information into proteins.
- miRNAs are able to turn off genes by inactivating mRNAs.
- A miRNA is complementary to a part of one or more mRNAs, that promotes cleavage or destroy them.
- **The miRNAs and their target-mRNAs have a many-to-many mirroring relationship**
→ We will use this property.

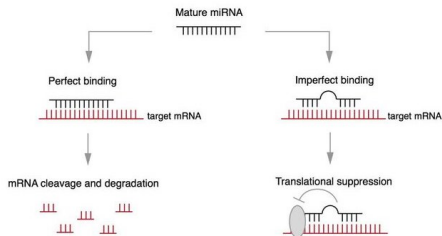


Figure: Mechanism of miRNA action. MiRNA can bind to specific regions of target mRNA transcripts and destabilizes the target transcript and/or blocks its translation. Source: [11].

Experiment and data

- *Striatum* of knock-in HD mice.
- Intervention on *polyQ* (CAG) length, one of $\{20, 80, 92, 111, 140, 175\}$.
- *Time* of evaluation of miRNA and mRNA expressions (log-fold change), either 2, 6 or 10 months.
- Results in $M = 13,616$ (mRNA) and $N = 1,143$ (miRNA) profiles (data points) in \mathbb{R}^{15} .

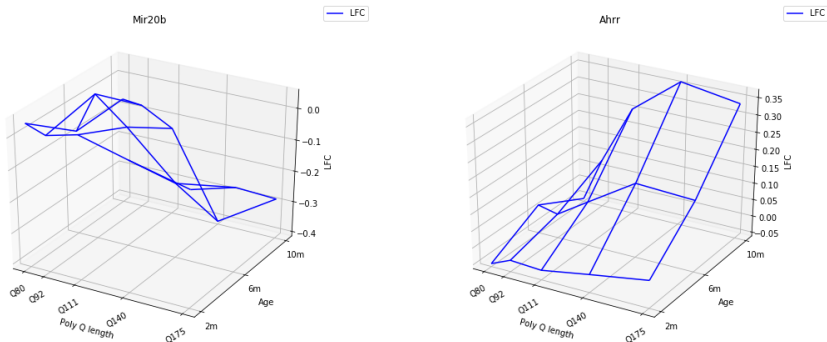


Figure: Left: profile y_n of a miRNA (Mir20b). Right: profile x_m of a mRNA (Ahrr). It is believed that Mir20b targets Ahrr.

Objective

Finding couples (miRNA, mRNA) that “collaborate”

Based on the profiles $\{x_1, \dots, x_M\}$ (mRNA profiles) and $\{y_1, \dots, y_N\}$ (miRNA profiles), we wish to identify collections $\{(x_m, y_n) : (m, n) \in \mathcal{S}\}$ gathering mRNAs and miRNAs that “collaborate”.

- An *ideal* match between a mRNA and a miRNA would consist of two profiles that display a *perfect mirroring relationship*: $y_n = -x_m$.
- We will relax this very strong biological hypothesis and consider loosened relationships $y \approx \theta(x)$ for a transformation $\theta \in \Theta$, where Θ is a parametric set containing $-id$.

- **Illustration:** profiles of two mRNA and miRNA which are believed to collaborate

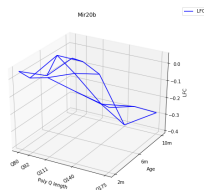


Figure: Profile of Mir20b (miRNA)

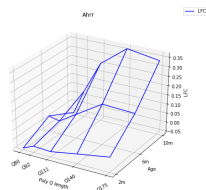


Figure: Profile of Ahrr (mRNA)

Proposal

- We develop a **procedure** called **WTOT-matching** to find collections $\{(x_m, y_n) : (m, n) \in S\}$ of mRNAs and miRNAs that “collaborate”.
- The procedure unfolds in **two steps**:
 - WTOT-...: consists in constructing a *similarity matrix* between mRNAs and miRNAs
 - we define the *similarity matrix* as an *optimal coupling matrix*;
 - we operationalize the search of **mirroring relationships**.
 - ...-matching: consists in deriving several sets of matched elements from the similarity matrix.

Modicum of optimal transport (1/2)

- Let $X := \{x_1, \dots, x_M\} \subset \mathbb{R}^d$ and $Y := \{y_1, \dots, y_N\} \subset \mathbb{R}^d$ be two data sets.
- For any $\omega \in \Omega_M := \{\omega \in (\mathbb{R}_+)^M : \|\omega\|_1 = 1\}$ and $\omega' \in \Omega_N$, define

$$\mu_X^\omega = \sum_{m \in \llbracket M \rrbracket} \omega_m \delta_{x_m}, \quad \nu_Y^{\omega'} = \sum_{n \in \llbracket N \rrbracket} \omega'_n \delta_{y_n}.$$

- The measures μ_X^ω and $\nu_Y^{\omega'}$ represent the two data sets.
 - Each x_m is given a weight ω_m .
 - Each y_n is given a weight ω'_n .
- The optimal transport (OT) matrix is defined as any element of

$$\arg \min_{P \in \Pi(\omega, \omega')} \langle C_{X,Y}, P \rangle_F,$$

where

- $\Pi(\omega, \omega')$ is the set of $P \in (\mathbb{R}_+)^{M \times N}$ such that $P \mathbf{1}_N = \omega$ and $P^\top \mathbf{1}_M = \omega'$;
 - $C_{X,Y} \in \mathbb{R}^{M \times N}$ is a **cost matrix** given by $(C_{X,Y})_{mn} := c(x_m, y_n)$ for some **cost function** $c : \mathbb{R}^d \times \mathbb{R}^d \rightarrow \mathbb{R}_+$;
 - $\langle C_{X,Y}, P \rangle_F := \sum_{(m,n) \in \llbracket M \rrbracket \times \llbracket N \rrbracket} (C_{X,Y})_{mn} P_{mn}$
- Computing the **arg min** is difficult and slow (and unicity is not guaranteed).

Modicum of optimal transport (2/2)

- Focus on **entropic-regularized OT**: for any $\gamma > 0$,

$$\mathcal{W}_\gamma(\mu_X^\omega, \nu_Y^{\omega'}) = \min_{P \in \Pi(\omega, \omega')} \{ \langle C_{X,Y}, P \rangle_F - \gamma E(P) \}$$

where $E(P) = - \sum_{(m,n) \in [M] \times [N]} P_{mn} (\log P_{mn} - 1)$. **Gain?**

- unique minimizer;
 - computing the **arg min** is much easier (Sinkhorn's algorithm).
- Introduce the **Sinkhorn loss**:

$$\bar{\mathcal{W}}_\gamma(\mu_X^\omega, \nu_Y^{\omega'}) := 2\mathcal{W}_\gamma(\mu_X^\omega, \nu_Y^{\omega'}) - \mathcal{W}_\gamma(\mu_X^\omega, \mu_X^\omega) - \mathcal{W}_\gamma(\nu_Y^{\omega'}, \nu_Y^{\omega'})$$

Gain?

- non-negative, symmetric, convex;
- metrizes convergence of measures;
- unbiased gradient estimates;
- interpolates between OT (its nice geometry) and Maximum Mean Discrepancy (its favorable high-dimensional sample complexity + sensitivity to differences in both location and shape of distributions).

WTOT-matching: WTOT-... (1/4)

- Introduce

$$\Theta := \left\{ \theta : \mathbb{R}^d \rightarrow \mathbb{R}^d, x \mapsto \theta(x) = \theta_1 x + \theta_2, \theta_1 \in T_1 \subset \mathbb{R}^{d \times d}, \theta_2 \in \mathbb{R}^d \right\},$$

where

- the matrices θ_1 are constrained;
 - in particular, their **diagonals** are made of negative values (\sim **mirroring relationship**);
 - $-\text{id} \in \Theta$.
- For all $\omega \in \Omega_M, \theta \in \Theta$, define

$$\mu_{\theta(X)}^{\omega} = \sum_{m \in [M]} \omega_m \delta_{\theta(X_m)}, \quad \nu_Y = \frac{1}{N} \sum_{n \in [N]} \delta_{Y_n}.$$

- Our **master program** is

$$\min_{\omega \in \Omega} \min_{\theta \in \Theta} \bar{W}_{\gamma} \left(\mu_{\theta(X)}^{\omega}, \nu_Y \right), \quad (\square)$$

where we are interested in the **minimizer** $(\hat{\omega}, \hat{\theta})$ and in the **optimal matrix** $\hat{P} \in \Pi(\hat{\omega}, N^{-1} \mathbf{1}_N)$ solving

$$\min_{P \in \Pi(\hat{\omega}, N^{-1} \mathbf{1}_N)} \left\{ \langle C_{\hat{\theta}(X), Y}, P \rangle_F - \gamma E(P) \right\}.$$

WTOT-matching: WTOT-... (2/4)

- We propose to solve (□) by iteratively updating ω and then θ .
- Given a kernel φ (standard normal density):
 - sample $\theta^{(0)}$ in Θ ;
 - iteratively for $0 \leq \tau < T$,
 1. define $\omega^{(\tau)} \in \Omega_M$ such that $\omega_m^{(\tau)} \propto \nu_Y \varphi\left(\frac{\cdot - \theta^{(\tau)}(x_m)}{h}\right)$ (all $m \in [M]$);
 2. solve $\theta^{(\tau+1)} \in \arg \min_{\theta \in \Theta} \bar{\mathcal{W}}_\gamma\left(\mu_{\theta(X)}^{\omega^{(\tau)}}, \nu_Y\right)$.
- Then we retrieve the corresponding OT matrix \hat{P} that solves

$$\mathcal{W}_\gamma\left(\mu_{\theta^{(T)}(X)}^{\omega^{(T)}}, \nu_Y\right) = \min_{P \in \Pi(\omega^{(T)}, N^{-1}\mathbf{1}_N)} \left\{ \langle C_{\theta^{(T)}(X), Y}, P \rangle_F - \gamma E(P) \right\}.$$

- Comments:
 - use of mini-batches in step 2;
 - $\theta^{(T)} : \mathbb{R}^d \rightarrow \mathbb{R}^d$ models to relax the mirroring relationships;
 - \hat{P}_{mn} can be interpreted as a similarity between x_m and y_n ;
 - $\omega^{(T)}$: weights to operationalize the many-to-many relationships.

WTOT-matching: ...-matching (3/4)

- Fix two integers $k, k' \geq 1$, let $\hat{\tau}$ be the quantile of order q of all the entries of \hat{P} .
- For every $m \in \llbracket M \rrbracket$ and $n \in \llbracket N \rrbracket$

$$\mathcal{N}_m^0 := \left\{ n \in \llbracket N \rrbracket : \hat{P}_{mn} \in \{\hat{P}_{m(1)}, \dots, \hat{P}_{m(k)}\} \text{ and } \hat{P}_{mn} \geq \hat{\tau} \right\},$$

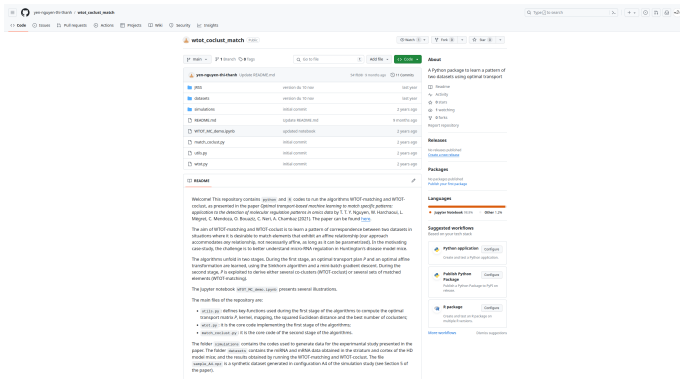
$$\mathcal{M}_n^0 := \left\{ m \in \llbracket M \rrbracket : \hat{P}_{mn} \in \{\hat{P}_{(1)n}, \dots, \hat{P}_{(k')n}\} \text{ and } \hat{P}_{mn} \geq \hat{\tau} \right\}.$$

- Define the most relevant matches

$$\mathcal{R} := \left\{ (m, n) \in \llbracket M \rrbracket \times \llbracket N \rrbracket : n \in \mathcal{N}_m^0 \text{ and } m \in \mathcal{M}_n^0 \right\}.$$

WTOT-matching: code (4/4)

- Code written in python and available on [this webpage](#).



- A **tutorial** is made available to show how simple it is to run the code.
- We adapt the Sinkhorn algorithm implemented by Aude Genevay and available [here](#).
- The stochastic gradient descents relies on the machine learning framework **pytorch**.

Real data application (1/4)

- We choose $k = k' = 10$, $q = 90\%$.
- Some facts:
 - we obtain 4234 non-empty \mathcal{N}_m s and 1043 non-empty \mathcal{M}_n s;
 - $\frac{\sum_{m \in \llbracket M \rrbracket} \text{card}(\mathcal{N}_m)}{\{m \in \llbracket M \rrbracket : \mathcal{N}_m \neq \emptyset\}} \approx 1.82$, $\frac{\sum_{n \in \llbracket N \rrbracket} \text{card}(\mathcal{M}_n)}{\{n \in \llbracket N \rrbracket : \mathcal{M}_n \neq \emptyset\}} \approx 6.04$.
- Our findings and their analysis are shared on [this website](#).

Optimal transport-based machine learning to match specific patterns: application to the detection of molecular regulation patterns in omics data

Updated on February 28, 2023

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Data availability
Full list of cell-to-cell pairs identified using optimal transport-based machine learning.

[Download](#) [View](#)

Data visualization
Availability of publications with:
• Megret et al., 2023 (MIRAMANT analysis) [PubMed link](#)
• Langfelder et al., 2018 (WGCNA-based analysis) [PubMed link](#)

[Link with associated publications](#) [Download](#) [Access](#)

Data annotation
Direct annotations of target mRNAs obtained by WTOT, MIRAMANT and WGCNA-based analyses.

[Annotations](#)

Code availability
The temporary WTOT source code can be found below.

[Link](#)

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Real data application: example of “monotonic” profiles (2/4)

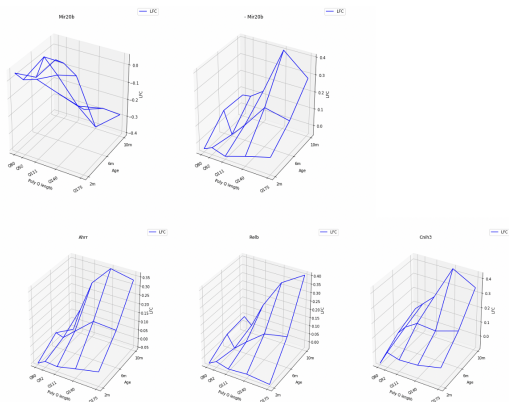


Figure: **Top:** profile y_n of Mir20b (left) and $-y_n$ (right). **Bottom:** profiles x_m ($m \in \mathcal{M}_n$) of the matched mRNAs Ahrr, Relb and Cnih3.

Real data application: example of “peaked” profiles (3/4)

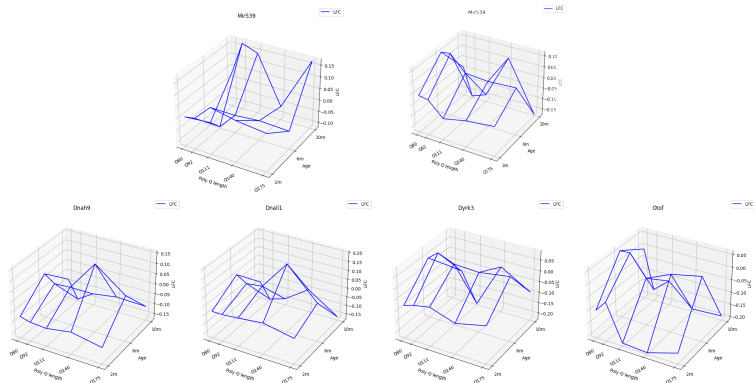


Figure: **Top:** profile y_n of Mir539 (left) and $-y_n$ (right). **Bottom:** profiles x_m ($m \in \mathcal{M}_n$) of the matched mRNAs Dnah9, Dnali1, Dyrk3, Otof.

Real data application: biological analysis of the results (4/4)

- A **biological analysis** is conducted to identify the more relevant pairs.
- A pair (x, y) is retained if and only if the mRNA whose profile is x and the miRNA whose profile is y are both among the 27,355 mRNAs and 1,478 miRNAs appearing in the TargetScan [5], MicroCosm [1] and miRDB [3] databases.
- The enrichment analysis reveals that the matchings output by **WTOT-matching** are
 1. primarily annotated for *extracellular matrix organization*, which relates to cell identity (due to the matchings labeled as neither peaked nor monotonic);
 2. secondarily annotated for *mitigation of host antiviral defense response* (due to the matchings labeled as monotonic), and for *conventional motile cilium* (due to the matchings labeled as peaked).

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Thanks for your attention!

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